Allogeneic Mesenchymal Stem Cells Infusion to an ALS Patient Proved to be Safe and Capable to Initiate Clinical Recuperation

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ABSTRACT

Introduction: No cure has been found for Amyotrophic Lateral Sclerosis (ALS) despite several studies using drugs or related compounds.

Background: Cellular, molecular and preclinical studies have suggested that the use of Mesenchymal Stem Cells (MSC) may represent a therapeutic option for ALS treatment. MSC exhibit a capability for self-renewal, differentiation (mesoderm and neuroectoderm phenotypes) and production/release of neuroprotective mediators. All these attributes epitomize an option for using MSC in ALS.

Materials and Methods: Minimally manipulated ex vivo expanded allogeneic bone marrow-derived MSC (0.6 x 106 MSC/kg body weight) were intrathecally infused to an ALS patient. Quantitative clinical assessments as well as vital signs, access site and neurological clinical assessment were monitored after cell infusion.

Results: Physical examination, vital signs, CBC and Chemistry Panel indicated no significant clinical effect (s) of the cell product infusion. Additionally, no evidence of adverse events/side effects was detected during a period of 18 months after cell infusion. Quantitative clinical assessments (forced vital capacity, hand held dynamometry and functional rating scale) at baseline and different time intervals after cell infusion revealed no evidence of acceleration, but a generalized delay in the progression of the disease.

Discussion: This data demonstrated that infusion of allogeneic MSC to an ALS patient was safe and feasible. Of main importance is the observation that even after 18 months of cell infusion, the patient has recognized a significant, stable and remarkable repossession of quality of life.

Conclusions: These results strongly suggest that the use of allogeneic MSC represents a safe and effective therapeutic option for ALS.

Introduction

Amyotrophic lateral sclerosis (ALS), commonly known as Lou Gehrig's disease, is a chronic progressive neurodegenerative disease that attacks motor neurons in the brain (upper motor neurons) and spinal cord (lower motor neurons). ALS affects over 350,000 of the world's population, and kills over 100,000 every year. It is commonly mentioned that ALS has no prejudice, since it occurs worldwide, with no age, ethnic or economic considerations. Despite several studies to assess effectiveness of drugs or related compounds on ALS and/ or related conditions, no cure has been found for ALS^{1,2}. Knowledge transpired from cellular, molecular and preclinical studies have put forward the notion that certain types of stem/progenitor cells may possibly contribute to repair and/or delay the disease-associated damage taking place in ALS or related conditions.

Mesenchymal stem cells (MSC) represent a type of adult stem cells that exhibit several features that epitomize an attractive option for treating central nervous system injuries, including ALS and traumatic spinal cord injury^{3,4}. Among these attributes the following are pertinent: a) capability for self-renewal, commitment and differentiation into several mesoderm and neuroectoderm phenotypes, b) production and release of a vast repertoire of "cell mediators" involved in the fine-tuning of distinct

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cellular microenvironments (neuroprotection), c) privileged immunophenotype capable of regulate immune responses and d) capability to manage oxidative stress⁵. The first report dealing with the use of adult stem cells in ALS patients, described that the dorsal spinal cord implantation of autologous bone marrow-derived MSC was safe, well tolerated and promising⁶. In a recent report, the same group emphasizes the necessity of better recruitment and selection of patients to test efficacy after the infusion of autologous MSC to ALS patients⁷.

Based on this information, as well as in additional evidence supporting the concept of a "cellular and molecular neurological trait" of MSC,

an IND was submitted and approved by the FDA, USA⁸. This phase I study was designed to assess the safety and feasibility of the intrathecal infusion of minimally manipulated ex vivo expanded autologous bone marrow-derived MSC, as shown by pre-clinical studies⁹.

DESCRIPTION OF THE CASE

The patient a 62-year-old Caucasian man was included in this study¹⁰ because of the progressive nature of ALS symptoms with no subjective or objective signs of decrease in the intensity of his muscular weakness or disability (Table 1, A and Figure 1, follow-up month 0).

Table 1. Pre- and post- cell infusion assessments.

A: Pre-infusion assessment	
Patient (DOB/gender)	March 1948; male
Firsts and reiterative symptoms (2003-2009); ALS Diagnosis and Medication (2010)	Limb weakness; started falling: strength weakness; speech changing to dysarthria/understandable; progressive weakness in legs/upper limb; muscle fasciculation. ALS diagnosed. Medication: Desvenlafaxine; Acetaminophen/Hydrocodone; Carisoprodol.
Baseline (Pre-infusion, August 2010)	Nerve examination: Cranial (olfactory, optic, oculomotor) and Trigeminal, Facial, Vestibule-cochlear and Accessory/Hypoglossal nerves were normal; Glossopharyngeal Vagus nerves were altered (phonation showed mild dysphonia and dysarthria). Motor examination: Dynamometry of uppers (average of 5 trials): right grip: 15 kg; left grip: 17.4 kg. Strength (MRC scale). Right: 4.2; Left: 4.1 (MRC scale). Wasting of first dorsal interosseous on both sides. Wide spread. Fasciculation in Calf Quads, Biceps, Deltoid and forearm muscles. Muscle tone appeared to be increased in legs more than in upper limbs. Deep tendon reflexes: increased on both sides with bilateral positive Babinski sign Sensory examination: Light touch, pin prick and vibration felt equally on both sides Cerebellar system: Finger nose test was unremarkable. Romberg negative. Gait: Difficult to get up from bed but manages to get up and sit down on the table; tends to be wide based and a tendency to sway due to muscular weakness; not able to perform tandem due to muscular weakness. Patient received the infusion of mesenchymal stem cells. For data related to neurological clinical examination (FVC, HHD, and ALS-FRS) at Baseline, see Figure 1.

B: Post-infusion assessment

For data related to neurological clinical examination (FVC, HHD, and ALS-FRS), see Figure 1.

2 months	Patient has 4/5 strength in neck flexors and extensors. Deltoids are 4/5 and biceps 4+/5. Wrist extensors are 3+ bilaterally. Finger extensors and grip are in psoas muscles and 3+. Strength 4/5 in the psoas muscles, quads and dorsiflexors. Hamstrings are about 3+. Deep tendon reflexes are symmetrical. Plantar responses are down going. Gait has a normal base. Station is mildly stooped. Stride out of balance and patient tends to wobble. Impression: ALS status post stem cells treatment is clinically stable.
6 months	Trace weakness in left deltoid. Good power in both upper extremities. Weakness: neck flexors (4+); iliopsoas (4); hamstrings (4); both quads (5) and anterior tibialis (4+).
12 months	No new problems and/or deterioration. Patient doing very well.
18 months (telephone assessment)	Evaluation pretty much the same as 6 months ago. He has a new tattoo on his left leg as part of his bucket list. Overall the patient seems to be doing fairly well and his ALS seems to be stable.

After fulfillment of the IND's inclusion/exclusion criteria¹¹, bone marrow aspiration was performed as the source of autologous MSC. The aspirate was sent to the GMP facility for isolation

and ex-vivo expansion of "native" MSC. Once in culture, the microscopic observation of vessel-attached cells revealed the presence of few MSC-like cells (CFU-F) and abundant cells with the distinction

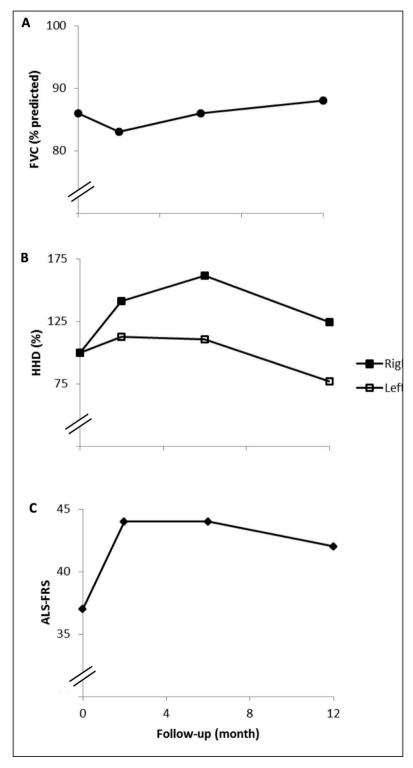


Figure 1. Neurological clinical examination at baseline and after cell infusion. *A*, FVC (forced vital capacity). *B*, HHD (hand held dynamometry). *C*, ALS-FRS (functional rating scale).

tive morphology of precursors/mature osteoclasts, consistent with previous observations¹². Since, after several medium changes, cultures contained mostly precursors/mature osteoclasts and non-proliferating MSCs (day 25 of 'extended' culture), the expansion attempt was discontinued. The patient was designated as a treatment failure and after 2 months, a second unsuccessful attempt to grow and expand bone marrow-derived MSC was initiated. According to protocol guidelines, the patient was withdrawn from the clinical study.

Based on the severity of ALS condition, the patient was treated with allogeneic MSC instead of autologous MSC, as the cell product to be infused. Allogeneic MSC, due to a distinctive immunophenotype, has been proven to be safe in animal and human studies. In the last case, allogeneic MSC have been safely transferred by diverse routes without eliciting immunologic reactions and/or side effects¹³.

STUDY OUTCOMES, PREPARATION AND INFUSION OF THE ALLOGENEIC CELL PRODUCT

Despite the caring and novel features of this study, the primary outcome was to assess the safety of the cell product infusion procedure, as assessed by absence of complications at the site of infusion and/or the appearance of new neurologic deficit/s not attributed to the natural progression of the disease. In turn, secondary outcomes were started to evaluate at different times after cell infusion, neurological changes in forced vital capacity (FVC), ALS functional rating scale scores (ALS-FRS) and hand held dynamometry (HHD).

Eligibility of a bone marrow donor was based on screening and testing for communicable diseases, as recommended (FDA; 21CFR 1271.75, 1271.80, 1271.85). For preparation of the donor cell product, bone marrow-derived MSC were *ex vivo* expanded following published procedures^{14,15}. Shortly after initiation of cell expansion, it was observed that MSC proliferated as the unique population of attached cells, which after two passages were released from the culture vessels, suspended in infusion medium (DPBS containing 0.05 % human serum albumin) and prepared for infusion ('cell product').

Aliquots of the cell product were prepared for lot release criteria tests (USP Sterility, Endotoxin, Mycoplasma, Viability and Gram's stain) and for immunophenotype identification. The expanded population of MSC expressed (≥ 98 %) antigens

CD73, CD90 and CD105 but not CD45 and CD34. After fulfillment of the lot-release criteria, the cell product was delivered for patient's infusion.

After sedation medication, the cell product (0.6 x 10⁶ MSC/kg body weight in 4 ml infusion media) was delivered to the lumbar intrathecal space (L3/4 level) by using a spinal needle under fluoroscopic guidance over a period of 6 minutes. After the infusion procedure, the patient was placed supine in Trendelenburg position for 1 h and then flat for another hour. Vital signs and access site were monitored for a period of 48 h. In turn, neurological clinical examination for a period up to 12 months was performed at TCA Cellular Therapy and at patient's County ALS Clinic (Our Lady of Lourdes Hospital, Lafayette, LA, USA).

RESULTS

Physical examination, vital signs, CBC and Chemistry Panel were performed at baseline and at week 2 and month 1, 6 and 12 post-cell infusion. The results of these evaluations indicated no significant clinical effect (s) after the infusion of the allogeneic stem cell product. Similarly, no adverse events or side effects were detected during a period of 12 months after cell infusion.

Results of neurological clinical examination at different time points after cell infusion are shown in Table 1, B and changes in forced vital capacity (FVC), hand held dynamometry (HHD) and ALS functional rating scale (ALS-FRS) are shown in Figure 1.

It is important to mention that in addition to the above evaluations, nerve conduction studies (lower and upper limbs) were also performed pre- and post- cell infusion (month 1, 2 and 6). As nerve conduction profiles at baseline revealed no significant alterations, alike results after cell infusion are not presented.

DISCUSSION AND CONCLUSIONS

The data here presented demonstrated that the intrathecal infusion of a cell product containing allogeneic *ex-vivo* expanded bone marrow-derived MSC to an ALS patient was feasible, safe and do not eliciting infusion-related adverse effects.

Infusion of the cell product was also effective as established by the clinical assessments performed shortly after infusion and during a 12 month follow-up period. As seen in Figure 1, Forced Vital Capacity (Figure 1 A) was kept rather constant during the entire follow up period. In turn, ALS-FRS score

(Figure 1 C) exhibited an early improvement which was maintained relatively constant during the 12 months follow up. In the case of HHD (%) values (Figure 1, B), shortly after stem cells infusion a transient improvement in right and left grip HHD was observed. While right grip values kept always higher than those at baseline, a time-dependent decreased of left grip values was observed.

All together, these data suggest that the infusion of allogeneic MSC produces in the patient a slow-down in disease progression as well as important survival benefits¹⁶⁻¹⁸.

Of main importance is the observation that even after 18 months of infusion of allogeneic MSC, the patient (telephone assessment) has recognized a significant, stable and remarkable repossession of quality of life.

Concurrently, the results presented in this study as well as those reported in a previous study showing functional deficiencies in MSC obtained from ALS patients¹⁹, strongly suggest that the infusion of allogeneic MSC represents a safe and effective alternative for the treatment of ALS.

The results described in this case report as well as in other studies published in the last two years²⁰⁻²², strongly put forward the notion that stem cells treatment could be a helpful therapeutic approach for ALS. Given the catastrophic nature of this disease and its great variability, these encouraging results embody not only a promising therapy but a strong reinforcement to "the principle of hope of patients that had no illusions of a panacea"²³.

We are aware that the biomedical community as well as ALS organizations and patients will follow with interest the progress of Clinical Protocol IND 13729²⁴, which provided the guidelines to threat the patient here described. This clinical study, which received from the FDA an Orphan Drug Designation (December 2010), was the first in the USA to utilize a non-invasive procedure to infuse adult and not laboratory differentiated stem cells to ALS patients.

CONSENT

Written informed consent was obtained from the patient for publication of this case report and for release of the Patient's Progress Report, as obtained by the patient from the County ALS Clinic (Our Lady of Lourdes Hospital, Lafayette, LA, USA). Copies of both consents are available for review by the Editor-in-Chief of the Journal.

CONFLICT OF INTERESTS

The Authors declare that they have no conflict of interests.

REFERENCES

- Shefner J, Cedarbaum JM, Cudkowicz ME, Maragakis N, Lee J, Jones D, Watson ML, Mahoney K, Chen M, Saikali K, Mao J, Russell AJ, Hansen RL, Malik F, Wolff AA. Safety, tolerability and pharmacodynamics of a skeletal muscle activator in amyotrophic lateral sclerosis. Amyotroph Lateral Scler 2012; 13(5): 430-438.
- Miller RG, Mitchell JD, Moore DH. Riluzole for amyotrophic lateral sclerosis (ALS)/motor neuron disease (MND). Cochrane Database Syst Rev 2012; 3: CD001447.
- 3. Vaquero J, Zurita M. Bone marrow stromal cells for spinal cord repair: a challenge for contemporary neurobiology. Histol Histopathol 2009; 24(1): 107-116.
- 4. Zhao CP, Zhang C, Zhou SN, Xie YM, Wang YH, Huang H, Shang YC, Li WY, Zhou C, Yu MJ, Feng SW. Human mesenchymal stromal cells ameliorate the phenotype of SOD1-G93A ALS mice. Cytotherapy 2007; 9(5): 414-426.
- 5. Minguell JJ, Allers C, Lasala GP. Mesenchymal stem cells and the treatment of conditions and diseases: the less glittering side of a conspicuous stem cell for basic research. Stem Cells Dev 2013; 22(2): 193-203.
- Mazzini L, Mareschi K, Ferrero I, Vassallo E, Oliveri G, Nasuelli N, Oggioni GD, Testa L, Fagioli F. Stem cell treatment in Amyotrophic Lateral Sclerosis. J Neurol Sci 2008; 265(1-2): 78-83.
- 7. Mazzini L, Mareschi K, Ferrero I, Miglioretti M, Stecco A, Servo S, Carriero A, Monaco F, Fagioli F. Mesenchymal stromal cell transplantation in amyotrophic lateral sclerosis: a long-term safety study. Cytotherapy 2012; 14(1): 56-60.
- Treatment of Amyotrophic Lateral Sclerosis, Phase I. FDA BB-IND 13729 (ClinicalTrials.gov Identifier: NCT01082653).
- Minguell JJ, Pereira A, Bartholomew P, Lasala GP. The Intrathecal Infusion of Mesenchymal Stem Cells into Healthy Rabbits is Safe and Devoid of Neurological or Clinical Complications. J Stem Cell Res Ther 2011; 01: 02.
- Treatment of Amyotrophic Lateral Sclerosis, Phase I. FDA BB-IND 13729 (ClinicalTrials.gov Identifier: NCT01082653).
- 11. Treatment of Amyotrophic Lateral Sclerosis, Phase I. FDA BB-IND 13729 (ClinicalTrials.gov Identifier: NCT01082653).
- 12. Allers C, Lasala G and Minguell JJ. Presence of Osteoclast Precursor Cells during ex vivo expansion of bone marrow-derived Mesenchymal Stem Cells for autologous use in cell therapy. Cytotherapy (in press), 2013.
- 13. Minguell JJ, Allers C, Lasala GP. Mesenchymal stem cells and the treatment of conditions and diseases: the less glittering side of a conspicuous stem cell for basic research. Stem Cells Dev 2013; 22(2): 193-203.
- 14. Conget PA, Minguell JJ. Phenotypical and functional properties of human bone marrow mesenchymal progenitor cells. J Cell Physiol 1999; 181(1): 67-73.
- Lasala GP, Silva JA, Minguell JJ. Therapeutic angiogenesis in patients with severe limb ischemia by transplantation of a combination stem cell product. J Thorac Cardiovasc Surg 2012; 144(2): 377-382.

- Czaplinski A, Yen AA, Appel SH. Forced vital capacity (FVC) as an indicator of survival and disease progression in an ALS clinic population. J Neurol Neurosurg Psychiatry 2006; 77(3): 390-392.
- Kaufmann P, Levy G, Thompson JL, Delbene ML, Battista V, Gordon PH, et al. The ALSFRSr predicts survival time in an ALS clinic population. Neurology 2005; 64: 38-43.
- Prabhakar S, Marwaha N, Lal V, Sharma RR, Rajan R, Khandelwal N. Autologous bone marrow-derived stem cells in amyotrophic lateral sclerosis: a pilot study. Neurol India 2012; 60: 465-469.
- Koh SH, Baik W, Noh MY, Cho GW, Kim HY, Kim KS, Kim SH. The functional deficiency of bone marrow mesenchymal stromal cells in ALS patients is proportional to disease progression rate. Exp Neurol 2012; 233(1): 472-480.

- 20. Prabhakar S, Marwaha N, Lal V, Sharma RR, Rajan R, Khandelwal N. Autologous bone marrow-derived stem cells in amyotrophic lateral sclerosis: a pilot study. Neurol India 2012; 60: 465-469.
- 21. Glass JD, Boulis NM, Johe K, Rutkove SB, Federici T, Polak M, Kelly C, Feldman EL. Lumbar intraspinal injection of neural stem cells in patients with amyotrophic lateral sclerosis: results of a phase I trial in 12 patients. Stem Cells 2012; 30(6): 1144-1151.
- 22. Ferraiuolo L, Frakes A, Kaspar BK. Neural stem cells as a therapeutic approach for amyotrophic lateral sclerosis. Mol Ther 2013; 21(3): 503-505.
- 23. Janson CG, Ramesh TM, During MJ, Leone P, Heywood J. Human intrathecal transplantation of peripheral blood stem cells in amyotrophic lateral sclerosis. J Hematother Stem Cell Res 2001; 10(6): 913-915.
- Treatment of Amyotrophic Lateral Sclerosis, Phase
 FDA BB-IND 13729 (ClinicalTrials.gov Identifier: NCT01082653).