Targeting Toll-Like Receptor 4: a promising strategy to prevent type 1 diabetes occurrence or recurrence

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Keywords: Type 1 diabetes, Islet transplantation, Prevention, Toll-like receptor, TLR4

Abstract

TLR4 is a transmembrane receptor of the innate immune system that recognize LPS of gramnegative bacteria. Its stimulation induces pro-inflammatory responses and also modulates adaptive immunity. In this article, we discuss the role of TLR4 in the activation and proliferation of T cells at the onset of autoimmune diabetes. We review the pathways involved in these observations. Finally, we show how targeting TLR4, by a variety of strategies, can prevent the occurrence of the disease but also its recurrence after allogeneic islet of Langerhans transplantation.

INTRODUCTION

Type 1 diabetes (T1D) is an autoimmune disease of unknown origin. It represents 5- 10% of the cases of diabetes diagnosed worldwide and mainly affects pediatric patients¹. It is characterized by autoimmune destruction - of the insulin-secreting cells (β -cells) of the islets of Langerhans. The decreased number of β -cells leads to an insufficient secretion of insulin and consequently to the dysregulation of glucose uptake and metabolism. There are genetic predispositions and environmental factors involved in the onset of the process leading to the destruction of the β -cells². In fact, there is growing evidence to suggest that T1D has remarkable interindividual heterogeneity in terms of clinical and immunopathological features³.

In this context, identifying modifiable factors is essential for the set-up of preventive strategies for patients at risk for T1D, but also to avoid recurrence of autoimmunity and allograft rejection after pancreas or islet transplantation and possibly, in the future, after autologous stem cell-derived islet transplantation or other innovative β -cell replacement or regeneration therapies. In this review, we will discuss the importance of chronic inflammation and insulitis in the pathogenesis of T1D. We will focus on the involvement of Toll-Like Receptor 4 (TLR4), a component of the inflammatory response, in the pathogenesis of T1D and on how it can be targeted for the prevention of the disease occurrence, or recurrence after β -cell replacement strategies.

TOLL-LIKE RECEPTORS AND AUTOIMMUNE DIABETES

The Toll-Like Receptor (TLR) family consists of 10 members in the human and 12 in the mouse⁴. They are either transmembrane molecules or expressed in intracellular compartments. Each transmembrane TLR is made of leucine-rich repeats (LRRs) at their N-terminal ends and a cytoplasmic Toll/IL-1 receptor (TIR) domain. TLRs are expressed in innate and adaptive immune cells, but also in fibroblasts, epithelial cells and adipocytes. Recently they have been identified in the β -cells^{5,6}. As pattern recognition receptors (PRRs), expressed by antigen-presenting cells (APC), they are able to recognize exogenous

pathogen-associated molecular patterns (PAMPs), that include bacteria-, virus-, fungus- and parasitederived components, but are also able to recognise endogenous damage-associated molecular patterns (DAMPs), such as self-DNA from necrotic or apoptotic cells. TLRs are essential to immune responses against pathogens. However inappropriate activation may promote inflammation and autoimmunity⁷. Recent discoveries have linked TLRs to inflammatory and autoimmune pathologies such as multiple sclerosis, autoimmune uveitis, arthritis, psoriasis, vasculitis, lupus erythematosus, inflammatory bowel disease and autoimmune diabetes⁸⁻¹².

The study of TLRs in autoimmune diabetes is complex. More than one cell, involved in the pathogenesis of T1D, express TLRs in addition to the multiple ligands that can interact with them. Thus, numerous contradictory results on the course of T1D have been published¹². TLR4 is a member of the TLR family. It complexes with myeloid differentiation factor 2 (MD-2) and recognizes bacterial lipopolysaccharide (LPS). Structural analysis revealed that five of six lipid chains of LPS bind to a hydrophobic pocket in MD-2, while the remaining chain binds to TLR4. TLR4-MD2-LPS complexes dimerize and induce signal transduction by recruiting adaptor proteins containing a TIR domain such as MyD88, TIRAP, TRAM and TRIF¹³. MyD88/ TIRAP-dependent pathway induces the production of pro-inflammatory cytokines whereas TRIF/ TRAM-dependent pathway is necessary for the production of type I IFN, a large subgroup of proteins that regulate the innate immune system to elicit responses against pathogens or tumors¹⁴. Although pathogen-recognition receptors in autoimmune diabetes have been widely studied, little is known about the role of TLR4 in the pathogenesis of this disease¹².

The expression of TLR4 in the islets of Langerhans has been documented to be deleterious both in murine and human β -cells^{6,15}. Exposure to LPS induces a loss of β -cells and decreases the synthesis and secretion of insulin^{5,6,10,15}. Moreover, it has been observed that the sera of type 1 diabetic patients exhibit increased levels of endogenous TLR4 ligands, such as endotoxins, HSP60 and HMGB1¹⁶. TLR4 expression is also upregulated on monocytes from type 1 diabetic patients¹⁷. Additionally, in a human cell model, CXCL10, through TLR4 signaling, induces β -cell death and dysfunction (18). Taken together these data provide convincing evidence for the deleterious effects on β -cells mediated by TLR4 activation.

TLR4 IN THE NON-OBESE DIABETIC (NOD) MOUSE MODEL

Little is known about the role of TLR4 in the development of autoimmune diabetes and contradictory observations are frequent. In an attempt to address this issue, a TLR4- deficient NOD mouse model has been developed¹⁹. Gülden et al¹² reported that TLR4-deficient NOD mice present an accelerated form of autoimmune diabetes12. The mean age of diabetes incidence was 17 weeks in TLR4-deficient NOD mice and 25 weeks in the control group. On the other hand, Devaraj et al¹⁹ described a protective phenotype in another TLR4 knock-out mouse model. Along a similar line, MyD88-deficient NOD mice, with impaired TLR4 signaling, do not develop diabetes²⁰. Kim et al⁷ also reported that the median age of diabetes onset was not affected by TLR4 gene knock-out. Moreover, the prevalence of diabetes was similar between TLR4+/+, TLR4+/and TLR4-/- mice¹².

The TLR4-deficient NOD mouse model is a useful tool to study the involvement of TLR4 in the pathogenesis of autoimmune diabetes. However, a general knockout of TLR4 appears to be not sufficient to understand the complex crosstalk between TLR4- expressing cells and to define their putative role in the onset of autoimmunity.

So far, the cell-specific role of TLR4 engagement has mainly been studied in β -cells and APC. The activation of TLR4 in murine β -cells impairs the insulin secretion and increases β -cell apoptosis in a murine model of type 2 diabetes²¹. On the other hand, TLR4 blockade preserves the survival of human and murine islets in the presence of LPS¹⁵. In this context, the engagement of TLR4 in β -cells may have a direct damaging effect.

TLR4 is mainly expressed in APC and its activation increases phagocytosis, antigen presentation and secretion of pro-inflammatory cytokines²². Macrophages can be found at the early stages of islet infiltration and probably play an important role in the onset of autoimmune diabetes^{23,24}. The activation of TLR4 in APC increases the secretion of pro-inflammatory cytokines in diabetic mice²¹. On the contrary, TLR4-deficient macrophages show reduced pro-inflammatory cytokine secretion and unresponsiveness to LPS exposition in NOD mice¹². On the basis of this evidence TLR4-deficient APC in NOD mice could have a possible protective phenotype against autoreactivity.

PREVENTION OF AUTOIMMUNE DIABETES BY TLR4 INHIBITION

Because of these considerations, we have explored whether TLR4 blockade could prevent the development of insulitis and the onset of autoimmune diabetes in NOD mice (25). Using CLI-095, a cyclohexene derivative that inhibits TLR4 signaling, we were able to observe a protective effect mediated by the decreased activation and proliferation of the diabetogenic autoreactive CD4+ T lymphocytes as well as IFN- γ production by these cells, both in the pancreatic lymph nodes and in the spleen, and consequently the inhibition of the autoimmune process. Interestingly, this was specifically observed in CD4, but not CD8 T-cells²⁵.

We further investigated the impact of TLR4 inhibition on CD4+ T lymphocytes in an adoptive transfer model of autoimmune diabetes. In this model, naïve NOD/BDC2.5 CD4+ T lymphocytes, expressing a rearranged TCR $\alpha\beta$, are transferred to immunocompromised NOD/rag1. CLI-095 treatment decreased the activation of CD4+ T lymphocytes and the number of IFN- γ -and IL-17A-producing CD4+ T lymphocytes, as compared to control. As observed in the NOD mouse model, TLR4 blockade impaired CD4+ T lymphocytes activation, proliferation and differentiation into diabetogenic effector cells, both in the pancreatic lymph nodes and in the spleen²⁵.

Since the injections of the CLI-095 TLR4 inhibitor were systemically (intraperitoneally) administered, numerous cells could have been targeted. This protective effect is potentially not only due to its effect on CD4+ T lymphocytes, but also on APCs. A protective effect on β -cells is also possible. Therefore, TLR4 inhibition could have the dual advantage of targeting the immune cells involved in the pathogenesis of T1D and directly increasing the viability of β -cells. Moreover, TLR4 is the receptor for inflammatory mediators (HMGB1, DAMPs, CXCL10), autoantigens (Heat-shock protein 60; HSP60) but also bacterial LPS, all being involved in the pathogenesis of T1D. Therefore, targeting inflammatory and autoimmune processes involved in the pathogenesis of T1D, using one single treatment could synergize the therapeutic efficiency and explain the promising results observed in NOD mice.

CLI-095 (also known as TAK-242) and Eritoran, are 2 inhibitors of TLR4 signaling, which have already been used in phase 3 clinical trials, in patients with severe sepsis^{26,27}. Regardless of the negative results of the trials, these TLR4 inhibitors were well tolerated. Therefore, inhibiting TLR4 by either of these agents in patients at risk for T1D could be a valid and feasible intervention. A derivative of HSP60 targeting TLR2 already demonstrated promising results on the preservation of β -cell function in newly diagnosed T1D patients²⁸. In addition, TLR polymorphisms are associated with susceptibility to infectious, inflammatory and autoimmune diseases²⁹. Because of the highly-conserved TLR functions throughout animal species, findings on the roles of TLRs in murine models could be easily translated to humans. Numerous clinical trials targeting TLR2,3,4,5,7,8 and TLR9 are ongoing for prevention and treatment of various cancers, allergic diseases, as well as inflammatory and autoimmune diseases³⁰.

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PREVENTION OF DIABETES RECURRENCE BY TLR4 blockade after islet transplantation

TLRs provide a well-recognized link between the innate and adaptive immune systems. In transplantation, they are a key link between ischemia-reperfusion injury and graft rejection³¹⁻³³. They play this role by enhancing antigen presentation, upregulating costimulatory molecule expression and stimulating proinflammatory cytokine and chemokine production. TLRs are typically engaged by exogenous ligands (such as LPS) in infection, but are also able to sense endogenous ligands, collectively known as damage-associated molecular patterns (DAMPs), such as key molecules involved in autoimmune diseases^{34,35} or necrotic cell products, usually released during ischemia-reperfusion injury in organ or cell transplantation^{31,34}.

TLRs were shown to play an important role in the inflammatory phenomena observed in the first stages of intraportal islet transplantation and leading to the early destruction of significant number of implanting endocrine cells. In particular, islet cells constitutively express TLRs, notably TLR2 and TLR4, and the isolation process upregulates TLR4 expression at their surface^{36,37}. Further, the engagement of islet-expressed TLRs by DAMPs at the time of transplantation leads to a massive release of proinflammatory cytokines and chemokines that contribute to the immediate destruction of a sizeable amount of implanting islet cells^{36,38}. The role of TLRs in these

processes was evidenced by experiments of syngeneic islet transplantation in TLR4^{-/-} murine models. Interestingly, absence of expression of TLR4 in the islet donors, not the recipients led to better engraftment, indicating that TLR4 expression by the islets, not the host innate immune system, was responsible for this phenomenon, which was largely mediated by the inhibition of NF-kappaB activation^{36,38}.

From another standpoint, TLR4-mediated dendritic cell (DC) activation can lead, via the nuclear factor kappa B pathway, to DC maturation and Treg suppression³⁹. In the setting of transplantation, ischemic injury can activate donor DCs by means of heat-shock protein70-TLR4 interaction. Donor-derived activated DCs can enhance immunogenicity of the graft, thus promoting graft rejection^{31,40}. Therefore, in addition to its impact on controlling autoimmune diabetes, TLR4 blockade could have the potential to mitigate the effects of islet or pancreas graft allogeneic immunogenicity.

There have been contradictory reports about the effect of TLR4 in allogeneic islet graft rejection, utilizing TLR4^{-/-} murine models. While all publications concur in finding a beneficial role of TLR4 inactivation on allogeneic islet graft rejection, significant differences are observed, depending on experimental design. Reports show indefinite islet graft survival in TLR4^{-/-} to wild type transplants, while others show the opposite (37,40,41). The different mouse strain combinations used may partly explain these discrepancies. They also exemplify the limitations of genetic manipulation and the differences between constitutional lack of expression and active blockade.

With an intent to develop an interventional strategy, we have explored the effect of a pharmacological TLR4 blockade, using monoclonal antibodies¹⁵. In mixed lymphocyte-islet reaction (MLIR) experiments, in which islets were co-cultured with allogeneic mononuclear cells, we were able to inhibit T-cell proliferation and IFN- γ expression with an anti-TLR4 monoclonal antibody (mAb). Interestingly, identical results were achieved in these in vitro experiments, using murine or human materials and a species-specific mAb¹⁵. In vivo, TLR4 blockade allowed indefinite allogeneic islet graft survival in a majority of animals. This effect was achieved only when both islet donors and islet graft recipients were treated with the mAb, indicated that TLR4 had to be inhibited both in the transplanted islet cells and the host immune system in order to achieve maximal effect¹⁵. In further mechanistic studies, animals tolerant to the islet graft (>100 days) were re-challenged with donor-specific or third party skin grafts. Donor-specific skin transplantation led to skin and islet graft loss within 12 days, whereas only skin got rejected, in the same time frame, after third party skin transplantation. This indicated that the protective effect of TLR4 blockade led to a phenomenon of graft accommodation rather than true immune tolerance¹⁵.

It should be noted here that, in these murine experiments, islets were transplanted under the kidney capsule, unlike the current practice in clinical islet transplantation, where islets are injected into the liver via the portal vein. A significant portion of transplanted islets are thought to be destroyed very early after injection by a phenomenon known as IBMIR ("Instant Blood-Mediated Inflammatory Reaction"), in which a cascade of inflammatory events is elicited by intravascular coagulation within the portal vein⁴². Involvement of TLRs in this phenomenon was reported, which makes it very likely that TLR4 blockade could enhance islet engraftment by controlling, or even inhibiting IBMIR⁴³.

TOLL-LIKE RECEPTOR BLOCKADE: TOWARD CLINICAL APPLICATION?

The literature reviewed in this article shows a clear potential for TLR -and in particular TLR4- blocking agents, both for the prevention of type 1 diabetes and also as a strategy aiming at promoting and prolonging islet graft survival. The mechanisms of action equally involve the targeting of inflammatory phenomena mediated by the innate immune system, but also purely adaptive immunity, such as occurs in autoimmunity and allogeneic graft rejection.

It is of interest that TLR4 blockade can be achieved in humans by a variety of agents, such as CLI-095 and Eritoran, both already tested in clinical trials^{26,27}, but also by human-specific monoclonal antibodies, which have been tested for another type of autoimmune disease (rheumatoid arthritis) in phase II studies⁴⁴. The availability of these compounds makes the clinical application of TLR4 targeting an immediate possibility for clinical trials aimed to prevent type 1 diabetes occurrence in subjects at risk and to enhance islet graft implantation and survival after pacreatic islet allotransplantation.

FUNDING:

This study was funded by a grant from the Swiss National Science Foundation (grant # 310030–149798).

CONFLICT OF INTEREST:

The authors have no conflict of interest to disclose.

References

- Maahs DM, West NA, Lawrence JM, Mayer-Davis EJ. Epidemiology of type 1 diabetes. Endocrinol Metab Clin North Am 2010; 39: 481-497.
- 2. Herold KC, Vignali DAA, Cooke A, Bluestone JA. Type 1 diabetes: translating mechanistic observations into effective clinical outcomes. Nat Rev Immunol 2013; 13: 243-256.
- Infante M, Ricordi C. Moving forward on the pathway of targeted immunotherapies for type 1 diabetes: the importance of disease heterogeneity. Eur Rev Med Pharmacol Sci 2019; 23: 8702-8704.
- 4. Kawasaki T, Kawai T. Toll-like receptor signaling pathways. Front Immunol 2014; 5: 461.
- 5. Li M, Song L, Gao X, Chang W, Qin X. Toll-like receptor 4 on islet β cells senses expression changes in high-mobility group box 1 and contributes to the initiation of type 1 diabetes. Exp Mol Med 2012; 44: 260-267.
- Garay-Malpartida HM, Mourão RF, Mantovani M, Santos IA, Sogayar MC, Goldberg AC. Toll-like receptor 4 (TLR4) expression in human and murine pancreatic beta-cells affects cell viability and insulin homeostasis. BMC Immunol 2011; 12: 18.
- Kim HS, Han MS, Chung KW, Kim S, Kim E, Kim MJ, Jang E, Lee HA, Youn J, Akira S, Lee MS. Toll-like receptor 2 senses beta-cell death and contributes to the initiation of autoimmune diabetes. Immunity 2007; 27: 321-333.
- Mills KHG. TLR-dependent T cell activation in autoimmunity. Nat Rev Immunol 2011; 11: 807-822.
- 9. Hong J, Leung E, Fraser AG, Merriman TR, Vishnu P, Krissansen GW. TLR2, TLR4 and TLR9 polymorphisms and Crohn's disease in a New Zealand Caucasian cohort. J Gastroenterol Hepatol 2007; 22: 1760-1766.
- Sacre SM, Andreakos E, Kiriakidis S, Amjadi P, Lundberg A, Giddins G, Feldmann M, Brennan F, Foxwell BM. The toll-like receptor adaptor proteins MyD88 and Mal/TIRAP contribute to the inflammatory and destructive processes in a human model of rheumatoid arthritis. Am J Pathol 2007; 170: 518-525.
- Abdollahi-Roodsaz S, Joosten LAB, Koenders MI, Devesa I, Roelofs MF, Radstake TR, Heuvelmans-Jacobs M, Akira S, Nicklin MJ, Ribeiro-Dias F, van den Berg WB. Stimulation of TLR2 and TLR4 differentially skews the balance of T cells in a mouse model of arthritis. J Clin Invest 2008; 118: 205-216.
- Gülden E, Ihira M, Ohashi A, Reinbeck AL, Freudenberg MA, Kolb H, Burkart V. Toll-like receptor 4 deficiency accelerates the development of insulin-deficient diabetes in non-obese diabetic mice. PLoS One 2013; 8: e75385.

- Park BS, Song DH, Kim HM, Choi B-S, Lee H, Lee JO. The structural basis of lipopolysaccharide recognition by the TLR4-MD-2 complex. Nature 2009; 458: 1191-1195.
- Yamamoto M, Sato S, Hemmi H, Uematsu S, Hoshino K, Kaisho T,Takeushi O, Takeda K, Akira S. TRAM is specifically involved in the Toll-like receptor 4–mediated MyD88- independent signaling pathway. Nat Immunol 2003; 4: 1144-1150.
- 15. Giovannoni L, Muller YD, Lacotte S, Parnaud G, Borot S, Meier RPH, Lavallard V, Bédat B, Toso C, Daubeuf B, Elson G, Shang L, Morel P, Kosco-Vilbois M, Bosco D, Berney T. Enhancement of Islet Engraftment and Achievement of Long-Term Islet Allograft Survival by Toll-Like Receptor 4 Blockade. Transplantation 2015; 99: 29-35.
- Devaraj S, Dasu MR, Park SH, Jialal I. Increased levels of ligands of Toll-like receptors 2 and 4 in type 1 diabetes. Diabetologia 2009; 52: 1665-1668.
- Devaraj S, Dasu MR, Rockwood J, Winter W, Griffen SC, Jialal I. Increased Toll-Like Receptor (TLR) 2 and TLR4 Expression in Monocytes from Patients with Type 1 Diabetes: Further Evidence of a Proinflammatory State. J Clin Endocrinol Metab 2008; 93: 578-583.
- Schulthess FT, Paroni F, Sauter NS, Shu L, Ribaux P, Haataja L, Strieter RM, Oberholzer J, King CC, Maedler K. CXCL10 Impairs β Cell Function and Viability in Diabetes through TLR4 Signaling. Cell Metab 2009; 9: 125-139.
- Devaraj S, Tobias P, Jialal I. Knockout of toll-like receptor-4 attenuates the pro-inflammatory state of diabetes. Cytokine 2011; 55: 441-445.
- 20. Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, Hu C, Wong FS, Szot G, Bluestone JA, Gordon JI, Chervonsky AV. Innate immunity and intestinal microbiota in the development of Type 1 diabetes. Nature 2008; 455: 1109-1113.
- Cucak H, Mayer C, Tonnesen M, Thomsen LH, Grunnet LG, Rosendahl A. Macrophage contact dependent and independent TLR4 mechanisms induce β-cell dysfunction and apoptosis in a mouse model of type 2 diabetes. PLoS One 2014; 9: e90685.
- 22. Takeda K, Kaisho T, Akira S. Toll-Like Receptors. Annu Rev Immunol 2003; 21: 335-376.
- Hanenberg H, Kolb-Bachofen V, Kantwerk-Funke G, Kolb H. Macrophage infiltration precedes and is a prerequisite for lymphocytic insulitis in pancreatic islets of pre-diabetic BB rats. Diabetologia 1989; 32: 126-134.
- Reddy S, Liu W, Elliott RB. Distribution of pancreatic macrophages preceding and during early insulitis in young NOD mice. Pancreas 1993; 8: 602-608.
- Alibashe-Ahmed M, Brioudes E, Reith W, Bosco D, Berney T. Toll-like receptor 4 inhibition prevents autoimmune diabetes in NOD mice. Sci Rep 2019; 9: 19350.
- 26. Opal SM, Laterre PF, Francois B, LaRosa SP, Angus DC, Mira JP, Wittebole X, Dugernier T, Perrotin D, Tidswell M, Jauregui L, Krell K, Pachl J, Takahashi T, Peckelsen C, Cordasco E, Chang CS, Oeyen S, Aikawa N, Maruyama T, Schein R, Kalil AC, Van Nuffelen M, Lynn M, Rossignol DP, Gogate J, Roberts MB, Wheeler JL, Vincent JL; ACCESS Study Group. Effect of Eritoran, an antagonist of MD2-TLR4, on mortality in patients with severe sepsis: The ACCESS Randomized Trial. JAMA 2013; 309: 1154-1162.

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- 27. Rice TW, Wheeler AP, Bernard GR, Vincent JL, Angus DC, Aikawa N, Demeyer I, Sainiti S, Amlot N, Cao C, Ii M, Matsuda H, Mouri K, Cohen J. A randomized, double-blind, placebo-controlled trial of TAK-242 for the treatment of severe sepsis. Crit Care Med 2010; 38: 1685-1694.
- 28. Raz I, Elias D, Avron A, Tamir M, Metzger M, Cohen IR. β-cell function in new- onset type 1 diabetes and immunomodulation with a heat-shock protein peptide (DiaPep277): a randomised, double-blind, phase II trial. Lancet 2001; 358: 1749-1753.
- Sutherland AM, Cook DN. Polymorphisms of the tolllike receptors and human disease. Clin Infect Dis 2005; 41: S403-S407.
- Hennessy EJ, Parker AE, O'Neill LA. Targeting Toll-like receptors: emerging therapeutics? Nat Rev Drug Discov 2010; 9: 293-307.
- Andrade CF, Waddell TK, Keshavjee S, Liu M. Innate immunity and organ transplantation: the potential role of toll-like receptors. Am J Transplant 2005; 5: 969-975.
- Jurewick M, Takakura A, Augello A, Mohavedi Naini S, Ichimura T, Zandi-Nejad K, Abdi R. Ischemic injury enhances dendritic cell immunogenicity via TLR4 and NF-kappa B activation. J Immunol 2010; 184: 2939-2948.
- Rahman AH, Taylor DK, Turka LA. The contribution of direct TLR signaling to T cell responses. Immunol Res 2009; 45: 25-36.
- Piccinini AM, Midwood KS. DAMPening inflammation by modulating TLR signaling. Mediators Inflamm 2010; 2010: pii672395.
- Shi H, Kokoeva MV, Inouye K, Tzameli I, Yin H, Flier JS. TLR4 links innate immunity and fatty acid-induced insulin resistance. J Clin Invest 2006; 116: 3015-3025.
- 36. Kruger B, Yin N, Zhang N, Yadav A, Coward W, Lal G, Zang W, Heeger PS, Bromberg JS, Murphy B, Schröppel B. Islet-expressed TLR2 and TLR4 sense injury and mediate early graft failure after transplantation. Eur J Immunol 2010; 40: 2914-2924.

- Goldberg A, Parolini M, Chin BY, Czismadia E, Otterbein LE, Bach FH, Wang H. Toll-like receptor 4 suppression leads to islet allograft survival. FASEB J 2007; 21: 2840-2848.
- 38. Gao Q, Ma LL, Gao X, Yan W, Williams P, Yin DP. TLR4 mediates early graft failure after intraportal islet transplantation. Am J Transplant 2010; 10: 1588-1596.
- 39. Garin A, Meyer-Hermann M, Contie M, Figge MT, Buatois V, Gunzer M, Toellner KM, Elson G, Kosco-Vilbois MH. Toll-like receptor 4 signaling by follicular dendritic cells is pivotal for germinal center onset and affinity maturation. Immunity 2010; 33: 84-95.
- 40. Ro H, Hong J, Kim BS, Lee EW, Kim MG, Han KH, Yeom HJ, Lee EM, Jeong JC, Oh KH, Ahn C, Yang J. Roles of Toll-like receptors in allogeneic islet transplantation. Transplantation 2012; 94: 1005-1012.
- Zhang N, Kruger B, Lal G, Luan Y, Yadav A, Zang W, Grimm M, Waaga-Gasser AM, Murphy B, Bromberg JS, Schröppel B. Inhibition of TLR4 signaling prolongs Treg- dependent murine islet allograft survival. Immunol Lett 2010; 127: 119-125.
- 42. Moberg L, Korsgren O, Nilsson B. Neutrophilic granulocytes are the predominant cell type infiltrating pancreatic islets in contact with ABO-compatible blood. Clin Exp Immunol 2005; 142: 125-131.
- 43. Vivot K, Langlois A, Jeandidier N, Bietiger W, Pinget M, Gies JP, Sigrist S. Instant blood-mediated inflammatory reaction during islet transplantation: the role of toll-like receptors signaling pathways. Transplant Proc 2011; 43: 3192-3194.
- 44. Monnet E, Choy EH, McInnes I, Kobakhidze T, de Graaf K, Jacqmin P, Lapeyre G, de Min C. Efficacy and safety of NI-0101, an anti-toll-like receptor 4 monoclonal antibody, in patients with rheumatoid arthritis after inadequate response to methotrexate: a phase II study. Ann Rheum Dis 2020; 79: 316-323.