

Mesenchymal stem cell-derived exosomes as drug delivery vehicles: an update

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ABSTRACT

Exosomes are extracellular vesicles with a diameter of 40-100 nm, which can be produced by several cells. As natural nanoscale vesicles, exosomes have the characteristics of low toxicity, low immunogenicity, strong homing ability, as well as the ability to carry several molecules, which make them potential drug delivery vehicles. Mesenchymal stem cells (MSCs) are an ideal source of exosomes because they can secrete and produce a large number of these vesicles. MSC-derived exosomes (MSCs-Exo) have regeneration and tissue repair functions similar to those of MSCs and can prevent the risk of graft versus host disease and infection caused by MSC transplantation. Therefore, MSCs-Exo are expected to become an effective substitute for the biological functions of MSCs. In this review, combined with the progress of preclinical and clinical studies, the potential therapeutic applications of MSCs-Exo in cardiovascular diseases, neurological disorders and malignancies was evaluated and discussed.

INTRODUCTION

Mesenchymal stem cells (MSCs) are multipotent stem cells with the ability of self-renewal and multidirectional differentiation. These cells are one of the most widely used stem cells and play an important role in regenerative medicine. MSCs arise from a wide range of sources and can be produced

by a variety of tissues, such as bone marrow, adipose tissue, muscle, placenta, umbilical cord, umbilical cord blood and dental pulp^{1,2}. MSCs have a variety of biological functions, including tissue repair, immunosuppression and neuroprotection^{3,4}. Accumulating evidence from preclinical and clinical studies indicated that the biological functions of MSCs are closely related to their paracrine actions, including their exosome-secreting ability^{5,6}.

Exosomes are extracellular vesicles secreted from cells under physiological and pathological conditions and can be used as an auxiliary tool for the diagnosis and treatment of several diseases⁷⁻⁹. Exosomes contain RNA molecules and proteins, which suggests that they can act as vehicles for bioactive compounds. Lai et al¹⁰ first identified exosomes derived from MSCs (MSCs-Exo) while they were isolating MSC medium. This study further showed that purified MSCs-Exo can play a similar role as that of MSCs in reducing the infarct size in a mouse model of myocardial ischemia/reperfusion (MI/R) injury¹⁰, thus providing a novel perspective for the study of mechanisms underlying the biological functions of MSCs. The secretion of MSC-Exo is similar to the secretion of exosomes derived from other sources. In particular, endosomes derive from plasma membrane invagination and then germinate inward to form nano-sized vesicles, resulting in multivesicular bodies that contain intraluminal vesicles. However, multivesicular bodies can have two different fates: i) one is to migrate towards the cell surface and fuse with the plasma membrane, releasing the exocrine body outside the cell by exocytosis; ii) the other is to fuse with lysosomes to degrade their content. Exo-



somes originate from the fusion between multivesicular bodies and the plasma membrane. In addition, MSCs-Exo have other characteristics in common with exosomes: they are composed of transmembrane proteins (CD9, CD63, CD81), integrin, major histocompatibility complex I and II molecules, heat shock proteins, polyvesicular synthesis related proteins, nucleic acids, cholesterol, glycerol phosphate and sphingomyelin^{11,12}. However, MSCs-Exo express specific markers, such as CD29, CD90, CD73 and CD44^{13,14}. Biological functions of MSCs-Exo are similar to those of MSCs. Moreover, MSCs-Exo reduce the risks associated with cell transplantation, are easily obtained, stored and used, have low immunogenicity and their production process is highly controllable¹⁵. These characteristics make MSCs-Exo potential natural drug delivery vehicles. Indeed, MSCs-Exo are being increasingly used in the treatment of cardiovascular diseases, neurological disorders and malignancies.

EXTRACTION AND IDENTIFICATION OF EXOSOMES

At present, the most commonly used method for exosome extraction is ultracentrifugation, which represents the “gold standard” for this procedure. However, this method can lead to exosome aggregation or contamination by other particles and cell fragments¹⁶. In addition, exosomes can be extracted by gradient density centrifugation, ultrafiltration centrifugation, molecular exclusion chromatography, polymer precipitation, immunoassay (magnetic beads, enzyme linked immunosorbent assay, flow cytometry and microfluidic techniques) and commercial kits, but there is still no ideal separation method to ensure content, purity and biological activity of exosomes at the same time^{17,18}. Since the extraction methods are based on the structure and size of exosomes and on the capture of some membrane proteins, it is difficult to distinguish exosomes from other vesicles and macromolecular protein complexes¹⁹. Therefore, exosome identification assay is necessary, including observation of exosome morphology under electron microscope, nano-tracking analysis of exosome size and quantity, and western blot analysis of exosome surface protein (CD9, CD63 and CD81) expression²⁰.

DRUG LOADING METHODS AND TYPES OF EXOSOMES

DIRECT DRUG LOADING

The drug is directly loaded into the purified exosome by co-incubation with the exosome, electro-

poration and chemical transfection. Small lipophilic molecules can be passively loaded into the exosome lipid bilayer membrane by co-incubating them with the exosomes, although the drug loading during this procedure is limited²¹. Electroporation means that repairable pores are instantly produced on the lipid bilayer membrane of the exosomes by applying a certain voltage by the electric field, thus introducing the drug into the exosomes. The parameters of electroporation are controllable; therefore, this procedure has wide application prospects⁹. However, some studies indicated that this method can induce the aggregation of exosomes, although the use of optimized parameters and pulse medium can effectively reduce the occurrence of this technical issue²². Chemical transfection usually employs commercial transfection reagents to load drugs into exosomes²³, but this method cannot completely separate drug-loaded exosomes from transfection agents. Thus, the potential toxicity of residual transfection reagents limits the application of this method.

INDIRECT DRUG LOADING

The drug is co-incubated or chemically transferred into the exosome-secreting cells. As the mechanism of drug delivery sorting into an exocrine action is more complex, it is difficult to control this process. Therefore, the practicability of this drug loading method is not strong. In addition, a new type of protein carrier which loads proteins into exosomes through optically reversible protein-protein interactions has been recently reported²⁴. This method allows to transfer most of the proteins into the cells, and also significantly improves the loading efficiency of exosomes.

TYPES OF EXOSOMES

Exosomes can carry bioactive molecules and chemical drugs, such as RNA (miRNA²⁵, lncRNA^{26,27}, siRNA²⁸), proteins²⁹, signaling molecules³⁰, paclitaxel³¹, curcumin³² and doxorubicin³³. MSCs-Exo not only act as drug delivery carriers, but also display a variety of biological functions similar to those of MSCs. Therefore, exosomes can also be used as “drugs”^{34,35}.

THE ADVANTAGES OF EXOSOMES AS DRUG DELIVERY VEHICLES

At present, liposome is a relatively mature drug vehicle, even though it has some limitations, such as the toxicity of synthetic liposome membrane and

low biocompatibility of targeted ligands³⁶. In the context of drug delivery carriers, exosomes display different advantages compared to synthetic liposomes, namely: 1) exosomes are extracellular vesicles actively secreted by cells, which can be regarded as natural liposomes and can overcome the limitations of synthetic liposomes mentioned above; 2) exosomes arise from the organism itself and have low immunogenicity³⁶; 3) exosomes can enter the blood circulation and the brain through the blood-brain barrier, and the intranasal administration allows the drug to quickly reach brain lesions, which opens up the possibility for non-invasive treatments of intracerebral diseases³⁸; 4) the particle size distribution of exosomes is 40-100 nm, which results in high permeability and retention effect at the site of solid tumors¹⁰; 5) exosomes have inherent homing and migration abilities and can be artificially manipulated to express specific molecules or improve their targeting ability³⁹⁻⁴¹.

CLINICAL APPLICATIONS OF MSCs-EXO AS DRUG DELIVERY VEHICLES

MSCs-Exo retain the physiological functions of MSCs. Therefore, MSCs-Exo can be used as “drugs”. In addition, MSCs-Exo can play a role as drug delivery vehicles to maximize drug effects.

CARDIOVASCULAR DISEASES

During the past decades, researchers have investigated the effects of MSCs-Exo in cardiovascular diseases. Emerging evidence indicated that MSCs-Exo can reduce the damage caused by MI/R through a variety of signaling pathways, such as the activation of Wnt/ β -catenin signaling and the induction of autophagy by AMPK/mTOR and Akt/mTOR pathways^{34,42}. In addition, MSCs-Exo can increase the levels of ATP and NADH in MI/R-injured heart, reduce the degree of oxidative stress and significantly reduce local and systemic inflammatory responses⁴³. In rat models of myocardial infarction, cardiac stem cells pre-treated with MSCs-Exo display higher survival rates and are able to promote the long-term recovery of cardiac function⁴⁴.

NEUROLOGICAL DISORDERS

Under the upsurge in MSCs-Exo use for the treatment of cardiovascular diseases, researchers also investigated the therapeutic role of MSCs-Exo in neurological disorders. In rat stroke models, MSCs-Exo can improve functional recovery, enhance neu-

rite remodeling, neurogenesis and angiogenesis⁴⁵. In rat models of acute ischemic stroke, MSCs-Exo can reduce the area of cerebral infarction and improve neurological function³⁷. Therefore, MSCs-Exo may represent a novel, promising strategy for the treatment of stroke. Some studies also showed that MSCs-Exo can down-regulate vascular endothelial growth factor-A (VEGF-A) to ameliorate blue light stimulation in retinal pigment epithelial cells and improve laser-induced retinal injury⁴⁶.

MALIGNANCIES

MSCs-Exo can also play an important therapeutic role in malignancies. At present, the main therapeutic approaches for treatment of solid tumors include surgical resection, chemotherapy, radiotherapy and hormone therapy. However, some of the abovementioned treatments have disadvantages, such as drug resistance and toxic effects on healthy tissues. Exosomes may overcome the shortcomings of the aforementioned approaches, displaying a high potential as targeted delivery carriers for solid tumors. Lee et al⁴⁷ demonstrated that MSCs-Exo can significantly down-regulate VEGF expression in breast cancer cells, inhibiting angiogenesis *in vitro* and *in vivo*. Lin et al⁴⁸ showed that MSCs-Exo can promote the migration of breast cancer cells through Wnt signal transduction pathway. Therefore, the exact mechanisms of action of MSCs-Exo in distinct tumor microenvironments need to be further investigated.

OTHER DISEASES

MSCs-Exo can repair osteochondral injury and promote a better recovery of steroid-induced early avascular necrosis of the femoral head by delivering mutant hypoxia inducible factor-1-alpha (HIF-1 α)⁴⁹. In a mouse model of acute liver failure induced by lipopolysaccharide/D-galactosamine, exosomes derived from human umbilical cord MSCs have been shown to repair the damaged liver tissue⁵⁰. In a mouse model of acute liver injury induced by carbon tetrachloride, the antioxidant and hepatoprotective effects of MSCs-Exo were superior than those of bifendate⁵¹. In a mouse model of autoimmune hepatitis, compared with MSCs-Exo, MSC-Exo containing miR-223 have been shown to significantly reduce serum levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST), as well the expression of pro-inflammatory cytokines at both protein and mRNA levels in hepatocytes⁵².

CONCLUSIONS

Exosomes are natural nanovesicles, which have several advantages as drug delivery vehicles. The functions of tissue repair and regeneration exerted by MSCs have been widely used for clinical purposes. MSCs-Exo exhibit both the aforementioned properties, and their biological characteristics can be altered by artificially modifying or interfering with the culture environment, which indicates that MSCs-Exo have wide clinical application prospects as delivery vehicles in the future. Although exosomes have several advantages and tend to be used in clinical research, we still lack a comprehensive understanding of the whole process of exosome production, transport and target-tissue uptake, which are all critical aspects that still need to be addressed carefully and cautiously.

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The authors declare that they have no conflict of interest to disclose.

REFERENCES

- Lanzoni G, Linetsky E, Correa D, Alvarez RA, Marttos A, Hirani K, Messinger Cayetano S, Castro JG, Paidas MJ, Efantis Potter J, Xu X, Glassberg M, Tan J, Patel AN, Goldstein G, Kenyon NS, Baidal D, Alejandro R, Vianna R, Ruiz P, Caplan AI, Ricordi C. Umbilical cord-derived mesenchymal stem cells for COVID-19 patients with acute respiratory distress syndrome (ARDS). *CellR4* 2020; 8: e2839.
- Jeon YJ, Kim J, Cho JH, Chung HM, Chae JI. comparative analysis of human mesenchymal stem cells derived from bone marrow, placenta, and adipose tissue as sources of cell therapy. *J Cell Biochem* 2016; 117: 1112-1125.
- Naji A, Eitoku M, Favier B, Deschaseaux F, Rouas-Freiss N, Sukanuma N. Biological functions of mesenchymal stem cells and clinical implications. *Cell Mol Life Sci* 2019; 76: 3323-3348.
- Wang X, Wang H, Lu J, Feng Z, Liu Z, Song H, Wang H, Zhou Y, Xu J. Erythropoietin-modified mesenchymal stem cells enhance anti-fibrosis efficacy in mouse liver fibrosis model. *Tissue Eng Regen Med* 2020 [publish online]. DOI: 10.1007/s13770-020-00276-2
- Xia W, Chen H, Xie C, Hou M. Long-noncoding RNA MALAT1 sponges microRNA-92a-3p to inhibit doxorubicin-induced cardiac senescence by targeting ATG4a. *Aging (Albany NY)* 2020; 12: 8241-8260.
- Ha DH, Kim HK, Lee J, Kwon HH, Park GH, Yang SH, Jung JY, Choi H, Lee JH, Sung S, Yi YW, Cho BS. Mesenchymal stem/stromal cell-derived exosomes for immunomodulatory therapeutics and skin regeneration. *Cells* 2020; 9: 1157.
- Garcia-Contreras M, Robbins PD. Exosomes and Microvesicles: Applications for Translational Research from Biomarkers to Therapeutic Applications-2013 ASMEV Meeting Report. *CellR4* 2013; 1: e412.
- Garcia-Contreras M, Ricordi C, Robbins PD, Oltra E. Exosomes in the pathogenesis, diagnosis and treatment of pancreatic diseases. *CellR4* 2014; 2: e807.
- Garcia-Contreras M, Messaggio F, Jimenez O, Mendez A. Differences in exosome content of human adipose tissue processed by non-enzymatic and enzymatic methods. *CellR4* 2015; 3: e1423.
- Lai RC, Arslan F, Lee MM, Sze NS, Choo A, Chen TS, Salto-Tellez M, Timmers L, Lee CN, El Oakley RM, Pasterkamp G, de Kleijn DP, Lim SK. Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. *Stem Cell Res* 2010; 4: 214-222.
- Alvarez-Viejo M. Mesenchymal stem cells from different sources and their derived exosomes: A pre-clinical perspective. *World J Stem Cells* 2020; 12: 100-109.
- Zhou X, Kalluri R. The biology and therapeutic potential of mesenchymal stem cells derived exosomes. *Cancer Sci* 2020 [publish online]. DOI: 10.1111/cas.14563
- Bousnaki M, Bakopoulou A, Kritis A, Koidis P. The Efficacy of stem cells secretome application in osteoarthritis: a systematic review of in vivo studies. *Stem Cell Rev Rep* 2020 [publish online]. DOI: 10.1007/s12015-020-09980-x
- Cooper LF, Ravindran S, Huang CC, Kang M. A Role for exosomes in craniofacial tissue engineering and regeneration. *Front Physiol* 2019; 10: 1569.
- Lou G, Chen Z, Zheng M, Liu Y. Mesenchymal stem cell-derived exosomes as a new therapeutic strategy for liver diseases. *Exp Mol Med* 2017; 49: e346.
- Lai RC, Yeo RW, Tan KH, Lim SK. Exosomes for drug delivery - a novel application for the mesenchymal stem cell. *Biotechnol Adv* 2013; 31: 543-551.
- Chen P, Ruan A, Zhou J, Huang L, Zhang X, Ma Y, Wang Q. Extraction and identification of synovial tissue-derived exosomes by different separation techniques. *J Orthop Surg Res* 2020; 15: 97.
- Wu X, Showiheen SAA, Sun AR, Crawford R, Xiao Y, Mao X, Prasadam I. Exosomes extraction and identification. *Methods Mol Biol* 2019; 2054: 81-91.
- Doyle LM, Wang MZ. Overview of extracellular vesicles, their origin, composition, purpose, and methods for exosome isolation and analysis. *Cells* 2019; 8: 727.
- Shamili FH, Bayegi HR, Salmasi Z, Sadri K, Mahmoudi M, Kalantari M, Ramezani M, Abnous K. Exosomes derived from TRAIL-engineered mesenchymal stem cells with effective anti-tumor activity in a mouse melanoma model. *Int J Pharm* 2018; 549: 218-229.
- Tian Y, Li S, Song J, Ji T, Zhu M, Anderson GJ, Wei J, Nie G. A doxorubicin delivery platform using engineered natural membrane vesicle exosomes for targeted tumor therapy. *Biomaterials* 2014; 35: 2383-2390.
- Hood JL, Scott MJ, Wickline SA. Maximizing exosome colloidal stability following electroporation. *Anal Biochem* 2014; 448: 41-49.

23. Wahlgren J, De LKT, Brisslert M, Vaziri Sani F, Telemo E, Sunnerhagen P, Valadi H. Plasma exosomes can deliver exogenous short interfering RNA to monocytes and lymphocytes. *Nucleic Acids Res* 2012; 40: e130.
24. Yim N, Choi C. Extracellular vesicles as novel carriers for therapeutic molecules. *BMB Rep* 2016; 49: 585-586.
25. Xu J, Bai S, Cao Y, Liu L, Fang Y, Du J, Luo L, Chen M, Shen B, Zhang Q. miRNA-221-3p in endothelial progenitor cell-derived exosomes accelerates skin wound healing in diabetic mice. *Diabetes Metab Syndr Obes* 2020; 13: 1259-1270.
26. Chen H, Xia W, Hou M. LncRNA-NEAT1 from the competing endogenous RNA network promotes cardioprotective efficacy of mesenchymal stem cell-derived exosomes induced by macrophage migration inhibitory factor via the miR-142-3p/FOXO1 signaling pathway. *Stem Cell Res Ther* 2020; 11: 31.
27. Xie F, Liu YL, Chen XY, Li Q, Zhong J, Dai BY, Shao XF, Wu GB. Role of MicroRNA, LncRNA, and exosomes in the progression of osteoarthritis: a review of recent literature. *Orthop Surg* 2020; 12: 708-716.
28. Shandilya S, Rani P, Onteru SK, Singh D. Natural ligand-receptor mediated loading of siRNA in milk derived exosomes. *J Biotechnol* 2020; 318: 1-9.
29. Thankam FG, Chandra I, Diaz C, Dilisio MF, Fleegel J, Gross RM, Agrawal DK. Matrix regeneration proteins in the hypoxia-triggered exosomes of shoulder tenocytes and adipose-derived mesenchymal stem cells. *Mol Cell Biochem* 2020; 465: 75-87.
30. Manini I, Ruaro ME, Sgarra R, Bartolini A, Caponnetto F, Ius T, Skrap M, Di Loreto C, Beltrami AP, Manfioletti G, Cesselli D. Semaphorin-7A on Exosomes: A Promigratory Signal in the Glioma Microenvironment. *Cancers (Basel)* 2019; 11: 758.
31. Salarpour S, Foroortanfar H, Pournamdari M, Ahmadi-Zeidabadi M, Esmaeeli M, Pardakhty A. Paclitaxel incorporated exosomes derived from glioblastoma cells: comparative study of two loading techniques. *Daru* 2019; 27: 533-539.
32. Wang H, Sui H, Zheng Y, Jiang Y, Shi Y, Liang J, Zhao L. Curcumin-primed exosomes potently ameliorate cognitive function in AD mice by inhibiting hyperphosphorylation of the Tau protein through the AKT/GSK-3beta pathway. *Nanoscale* 2019; 11: 7481-7496.
33. Li D, Yao S, Zhou Z, Shi J, Huang Z, Wu Z. Hyaluronan decoration of milk exosomes directs tumor-specific delivery of doxorubicin. *Carbohydr Res* 2020; 493: 108032.
34. Cui X, He Z, Liang Z, Chen Z, Wang H, Zhang J. Exosomes from adipose-derived mesenchymal stem cells protect the myocardium against ischemia/reperfusion injury through wnt/beta-catenin signaling pathway. *J Cardiovasc Pharmacol* 2017; 70: 225-231.
35. Kong LY, Liang MY, Liu JP, Lai P, Ye JS, Zhang ZX, Du ZM, Yu JJ, Gu L, Xie FC, Tang ZX, Liu ZY. Mesenchymal stem cell-derived exosomes rescue oxygen-glucose deprivation-induced injury in endothelial cells. *Curr Neurovasc Res* 2020 [publish online]. DOI: 10.2174/1567202617666200214103950
36. Sercombe L, Veerati T, Moheimani F, Wu SY, Sood AK, Hua S. Advances and Challenges of Liposome Assisted Drug Delivery. *Front Pharmacol* 2015; 6: 286.
37. Chen KH, Chen CH, Wallace CG, Yuen CM, Kao GS, Chen YL, Shao PL, Chen YL, Chai HT, Lin KC, Liu CF, Chang HW, Lee MS, Yip HK. Intravenous administration of xenogenic adipose-derived mesenchymal stem cells (ADMSC) and ADMSC-derived exosomes markedly reduced brain infarct volume and preserved neurological function in rat after acute ischemic stroke. *Oncotarget* 2016; 7: 74537-74556.
38. Long Q, Upadhy D, Hattiangady B, Kim DK, An SY, Shuai B, Prockop DJ, Shetty AK. Intranasal MSC-derived A1-exosomes ease inflammation, and prevent abnormal neurogenesis and memory dysfunction after status epilepticus. *Proc Natl Acad Sci U S A* 2017; 114: E3536-E3545.
39. Altanerova U, Jakubecova J, Benejova K, Priscakova P, Pesta M, Pitule P, Topolcan O, Kausitz J, Zduriencikova M, Repiska V, Altaner C. Prodrug suicide gene therapy for cancer targeted intracellular by mesenchymal stem cell exosomes. *Int J Cancer* 2019; 144: 897-908.
40. Li H, Liu D, Li C, Zhou S, Tian D, Xiao D, Zhang H, Gao F, Huang J. Exosomes secreted from mutant-HIF-1alpha-modified bone-marrow-derived mesenchymal stem cells attenuate early steroid-induced avascular necrosis of femoral head in rabbit. *Cell Biol Int* 2017; 41: 1379-1390.
41. Wang X, Chen Y, Zhao Z, Meng Q, Yu Y, Sun J, Yang Z, Chen Y, Li J, Ma T, Liu H, Li Z, Yang J, Shen Z. Engineered exosomes with ischemic myocardium-targeting peptide for targeted therapy in myocardial infarction. *J Am Heart Assoc* 2018; 7: e008737.
42. Liu L, Jin X, Hu CF, Li R, Zhou Z, Shen CX. Exosomes derived from mesenchymal stem cells rescue myocardial ischaemia/reperfusion injury by inducing cardiomyocyte autophagy via AMPK and Akt pathways. *Cell Physiol Biochem* 2017; 43: 52-68.
43. Arslan F, Lai RC, Smeets MB, Akeroyd L, Choo A, Aguor EN, Timmers L, van Rijen HV, Doevendans PA, Pasterkamp G, Lim SK, de Kleijn DP. Mesenchymal stem cell-derived exosomes increase ATP levels, decrease oxidative stress and activate PI3K/Akt pathway to enhance myocardial viability and prevent adverse remodeling after myocardial ischemia/reperfusion injury. *Stem Cell Res* 2013; 10: 301-312.
44. Zhang Z, Yang J, Yan W, Li Y, Shen Z, Asahara T. Pretreatment of cardiac stem cells with exosomes derived from mesenchymal stem cells enhances myocardial repair. *J Am Heart Assoc* 2016; 5: e002856.
45. Hong SB, Yang H, Manaenko A, Lu J, Mei Q, Hu Q. Potential of exosomes for the treatment of stroke. *Cell Transplant* 2019; 28: 662-670.
46. He GH, Zhang W, Ma YX, Yang J, Chen L, Song J, Chen S. Mesenchymal stem cells-derived exosomes ameliorate blue light stimulation in retinal pigment epithelium cells and retinal laser injury by VEGF-dependent mechanism. *Int J Ophthalmol* 2018; 11: 559-566.
47. Lee JK, Park SR, Jung BK, Jeon YK, Lee YS, Kim MK, Kim YG, Jang JY, Kim CW. Exosomes derived from mesenchymal stem cells suppress angiogenesis by down-regulating VEGF expression in breast cancer cells. *PLoS One* 2013; 8: e84256.

48. Lin R, Wang S, Zhao RC. Exosomes from human adipose-derived mesenchymal stem cells promote migration through Wnt signaling pathway in a breast cancer cell model. *Mol Cell Biochem* 2013; 383: 13-20.
49. Zhang S, Chu WC, Lai RC, Lim SK, Hui JH, Toh WS. Exosomes derived from human embryonic mesenchymal stem cells promote osteochondral regeneration. *Osteoarthritis Cartilage* 2016; 24: 2135-2140.
50. Jiang L, Zhang S, Hu H, Yang J, Wang X, Ma Y, Jiang J, Wang J, Zhong L, Chen M, Wang H, Hou Y, Zhu R, Zhang Q. Exosomes derived from human umbilical cord mesenchymal stem cells alleviate acute liver failure by reducing the activity of the NLRP3 inflammasome in macrophages. *Biochem Biophys Res Commun* 2019; 508: 735-741.
51. Jiang W, Tan Y, Cai M, Zhao T, Mao F, Zhang X, Xu W, Yan Z, Qian H, Yan Y. Human umbilical cord MSC-derived exosomes suppress the development of CCl₄-induced liver injury through antioxidant effect. *Stem Cells Int* 2018; 2018: 6079642.
52. Chen L, Lu FB, Chen DZ, Wu JL, Hu ED, Xu LM, Zheng MH, Li H, Huang Y, Jin XY, Gong YW, Lin Z, Wang XD, Chen YP. BMSCs-derived miR-223-containing exosomes contribute to liver protection in experimental autoimmune hepatitis. *Mol Immunol* 2018; 93: 38-46.